

## MONITORING OF HEAVY METALS CONTENT IN FISH MUSCLE TISSUE ALONG THE SAVA RIVER IN SERBIA

Radmila MARKOVIĆ<sup>1</sup>, Svetlana GRDOVIĆ<sup>1</sup>, Jelena JANJIC<sup>2\*</sup>,  
Dragoljub JOVANOVIĆ<sup>1</sup>, Dejan PERIĆ<sup>1</sup>, Vladimir DIMITRIJEVIĆ<sup>3</sup>,  
Stamen RADULOVIC<sup>1</sup>, Dragan ŠEFER<sup>1</sup>

<sup>1</sup>University of Belgrade, Faculty of Veterinary Medicine, Department for Animal Nutrition and Botany, Bulevar Oslobođenja 18, 11000 Belgrade, Serbia; <sup>2</sup>University of Belgrade, Faculty of Veterinary Medicine, Department for Economics and statistics, Bulevar Oslobođenja 18, 11000 Belgrade, Serbia; <sup>3</sup>University of Belgrade, Faculty of Veterinary Medicine, Department for Animal Breeding, Bulevar Oslobođenja 18, 11000 Belgrade, Serbia.

(Received 03 October, Accepted 28 November 2025)

The aim of this study was to examine the presence of heavy metals (Pb, Cd and Hg) in the muscle tissue of fish from the Sava River, from two locations: upstream of Belgrade near Obrenovac and in Belgrade. In order to determine the content of heavy metals in fish muscle tissue, 196 samples of different types of fish were collected: Prussian carp, carp, perch and catfish, during the fall and spring of 2023 – 2025. The content of heavy metals was determined by means of atomic absorption spectrometry. The highest average content of Pb ( $0.282 \pm 0.068$  mg/kg) and Hg ( $0.303 \pm 0.116$  mg/kg) was found in the muscle tissue of catfish, while the lowest average content of Pb and Hg was determined in the muscle tissue of perch ( $0.088 \pm 0.025$  mg/kg), ( $0.090 \pm 0.038$  mg/kg), respectively. The highest average content of Cd was recorded in the muscle tissue of carp ( $0.103 \pm 0.030$  mg/kg), while the lowest was found in the muscle tissue of catfish ( $0.032 \pm 0.015$  mg/kg). In all tested samples, the determined concentrations of Pb and Hg were below the maximum allowed concentrations regulated by the Rulebook of the Republic of Serbia and European Union regulations. The determined concentrations of Cd in the muscle tissue of carp, catfish and Prussian carp were on several locations above the maximum allowed concentration of 0.05 mg/kg of fresh fish. Fish from this part of the Sava river cannot be considered completely safe for human consumption, especially when it comes to species that show a tendency to accumulate Cd. Overall, seasonal variations were more evident than spatial differences, with several species showing moderately higher metal concentrations in spring, particularly in 2025, indicating subtle but recurring seasonal tendencies. Differences between the territories upstream of Belgrade and within Belgrade were generally small, although mercury levels were consistently higher at the downstream location, suggesting a mild but stable spatial influence.

**Keywords:** heavy metals, fish, environmental pollution, meat quality, nutrition

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\*Corresponding author: e-mail: jelena.janjic@vet.bg.ac.rs

## INTRODUCTION

Environmental pollution today is increasingly pronounced due to various biological and anthropological influences such as volcanic activity, soil and rock erosion, industrial development, chemicalization of agriculture, burning of fossil fuels, urbanization, as well as due to the application of inadequate environmental protection measures [1,2]. These are some of the main factors of pollution and presence of various contaminants in the ecosystem. The main environmental contaminants are various organic pollutants (Persistent Organic Pollutants – POP's), which include polycyclic aromatic hydrocarbons (PAH), polychlorinated biphenyls (PCB), organochlorine pesticides (OCP), dioxins [3], and heavy metals such as lead (Pb), cadmium (Cd) and mercury (Hg) are among the most important industrial pollutants [4,5]. Contamination with these metals is present in air, soil, sediment, plants and water. In the water environment, metals are found in low quantities, but due to the mentioned sources of contamination, their concentrations in the waters have increased [6]. When heavy metals infiltrate into water systems, they are dissolved, stored in sediments and cannot be degraded [7]. Therefore, their occurrence in sediments, water courses and living organisms indicates the existence of natural or anthropogenic sources of pollution [8,9]. As the majority of industrial wastewater in Serbia is discharged without adequate treatment [10], heavy metals released in this way will accumulate in aquatic organisms, including fish [11].

Bioaccumulation of heavy metals in fish tissues has been described in many studies, because they are at the top of the food chain in aquatic systems [5,12]. Heavy metals can accumulate in fish tissues through feed, gills, skin, and water intake [13]. The type of feed also has an important influence on the accumulation of heavy metals in fish tissues [14,15]. Thus, carnivorous fish, due to biomagnification, accumulate larger amounts of metals compared to species that are at a lower trophic level [16]. On the other hand, for bottom-feeding fish, sediment is the primary source of heavy metals [17]. The degree of bioaccumulation of metals in fish tissues is influenced by other factors such as body weight, age, sex, species and physiological state of the fish, calcium and organic ligand concentrations [18], as well as water temperature and pH value of the medium [19,20]. Also, the accumulation of heavy metals in all fish tissues does not have the same efficiency, so the muscle tissue of fish contains smaller amounts of heavy metals compared to the internal organs, liver, kidneys and gills [21].

As fish is an important food in the human diet, its consumption is one of the main sources of human exposure to heavy metals [12,22]. The assessment of the presence of heavy metals in fish muscle tissue is very important, so that they do not pose a danger to human health and that the concentrations are below the permitted levels [23]. The heavy metals, that is: lead (Pb), cadmium (Cd) and mercury (Hg) have no physiological role, persist in the ecosystem for a long time, and are toxic when present in traces [6,24]. They are known as non-essential elements and if ingested over a long period of time can show harmful effects for all organisms [25]. Many studies have described

that high concentrations of mercury in fish negatively affect its cardioprotective effect, and its presence can lead to neurological diseases in humans [26,27]. Cadmium and lead have an extremely negative effect on the health of children and adults. Lead accumulates in soft tissues and can cause liver, kidney and brain damage. Cadmium can lead to anemia, hypertension and prostate cancer [28]. Although the benefits of consuming fish are significant (quality proteins, the presence of polyunsaturated fatty acids, vitamins and minerals) [29,30], the presence of heavy metals in fish meat can indicate the harmfulness of consuming this food for human health. As the Sava River is important for fishing [16,31], caught fish can pose a significant risk to human health [32].

The aim of this research was to examine the content of heavy metals (Pb, Cd and Hg) in the muscle tissue of four fish species from the Sava River. The determined content of heavy metals in fish meat, indicates the pollution of the environment with harmful metals, which are found in the sediment and in the water, and consequently also in various fish tissues. An even bigger problem is that the Sava River flows through several European countries, where it can be polluted by various pollutants, and the level of contamination can even further increase in Serbia.

The content of the before mentioned toxic metals is regulated by National and European Regulations, that are harmonized, and is defined as the maximum allowed concentration in fish meat.

## **MATERIALS AND METHODS**

### **Sampling and study area**

For this study, fish were caught from the Sava River from two different locations. The first location is located 25 km upstream from Belgrade, Serbia, near the town of Obrenovac (44°44'49.3"N;19°47'24.5"E) (sampling point A), and the other is downstream, in Belgrade, before the confluence of the Sava River and the Danube River (44°47'48.1"N;20°24'43.4"E) (sampling point B), in order to observe the levels of heavy metals from the perspective of big city pollutants. The determination of the content of heavy metals was carried out on the following types of fish: Prussian Carp (*Carassius auratus gibelio*), carp (*Cyprinus carpio*), catfish (*Silurus glanis*) and perch (*Sander lucioperca*). All types of fish were collected from professional fishermen in four periods, during the fall and spring of 2023 – 2025. A total of 192 muscle samples of four fish species were analyzed, six samples of each species from the Sava River from two locations in four periods. After harvesting, each fish was identified to species level and then euthanized with 3-aminobenzoic acid ethyl ester (MS-222). The fish were then transported to the laboratory in a cooling box (hand-held refrigerator) at + 4 °C. The fish were dissected in the laboratory, and muscle samples (about 100 g) were taken from under the dorsal fin and transferred to polyethylene containers that were

previously cleaned (10%) with nitric acid and washed three times with deionized water. The packed samples were then frozen and stored at  $-20^{\circ}\text{C}$  until heavy metal analysis was performed. Fish muscle was chosen as the target tissue in order to determine the risk that metal pollution poses to human health, and at the same time it is an indicator of the state of the ecosystem.

## Reagents

For the determination of the content of the tested metals (Pb, Cd and Hg), the chemicals used to digest the samples, nitric acid,  $\text{HNO}_3$  (65%) and hydrogen peroxide,  $\text{H}_2\text{O}_2$  (30%), were of high purity grade (J.T. Baker). Standard solutions of elements, which were used for calibration, were prepared by diluting basic solutions (J.T. Baker) with a concentration of  $1000\text{ mg L}^{-1}$ . Deionized water (resistivity  $18.2\text{ M}\Omega\text{cm}^{-1}$ ) obtained from a Millipore Elix UV 10 system was used for all dilutions.

## Apparatus

Samples preparation was performed using a MW 3000 microwave closed system (Anton Paar GmbH, Graz, Austria) with a MF 100 cuvette. The content of all elements was determined on an Analyst 700 atomic absorption spectrometer with an MHS system (Perkin Elmer, Shelton, USA). Concentrations of elements (Pb and Cd) were detected by the graphite technique (GFAAS-800) using an auto sampler on the Analyst 700 apparatus (Perkin Elmer), while the mercury content was determined by the cold vapor technique (CVAAS) with a discontinuous system (MHS-15) on the same apparatus. The operating parameters of the atomic absorption spectrometer (Analyst 700) for the determination of heavy metals were adjusted according to the manufacturer's recommendations (Perkin Elmer) and are shown in Tables 1 and 2.

**Table 1.** Instrumental parameters for graphite furnace (GFAAS) using Analyst 700

Metals	Wavelength (nm)	Parameters	Step 1 (Drying)	Step 2 (Pyrolysis)	Step 3 (Atomization)	Step 4 (Cleaning)
Pb	283.3	Temperature ( $^{\circ}\text{C}$ )	130	450	1900	2500
		Ramp time (s)	10	15	0	0
		Hold time (s)	30	10	4	5
Cd	228.8	Temperature ( $^{\circ}\text{C}$ )	130	350	1200	2500
		Ramp time (s)	10	15	0	0
		Hold time (s)	30	10	4	5

**Table 2.** Instrumental parameters for cold vapor (CVAAS) using Analyst 700

Metals	Wavelength (nm)	Acetylene flow (L min <sup>-1</sup> )	Air flow (L min <sup>-1</sup> )	Slit (nm)	N <sub>2</sub> pressure (bar)
Hg	253.7	/	12.0	0.2	4.0-5.0

### Heavy metals analysis

Fish muscle tissue samples were thawed at room temperature, then mechanically ground and homogenized. To determine the content of the tested metals, about  $1 \pm 0.001$  g of the sample was weighed on an analytical balance in cuvettes for sample preparation. The samples were treated with a mixture of HNO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub> (7:1, v/v) and performed to microwave digestion for 30 minutes. The digestion process was carried out with a program for decomposing fish meat samples, with a range of 250 to 630 W, in four steps, for 30 minutes. After cooling to room temperature, the prepared samples were quantitatively transferred into a measuring vessel, which was filled up to the marked level with deionized water. After the preparation, the content of toxic elements (Pb, Cd and Hg) was determined on the Analyst 700 apparatus. According to the manufacturer's instructions for the graphite technique (GFAAS-800), prescribed modifiers were used. To determine the content of Pb, the modifier primary ammonium phosphate was used, while for Cd, a mixture of magnesium nitrate and palladium chloride was used. The Hg content was determined using a hydride discontinuous system (MHS-15) after reduction with sodium boron hydride (NaBH<sub>4</sub>). All prepared samples were analyzed twice for each heavy metal. The content of each element (Pb, Cd and Hg) was measured in the peak area mode against a calibration curve, taking into account the masses of the samples and possibly applied dilutions. Metal concentrations are expressed in milligrams per kilogram of fresh fish (w/w). The detection limits of heavy metals in fresh fish meat were 0.02, 0.005 and 0.0125 mg kg<sup>-1</sup> for Pb, Cd and Hg, respectively. The quality of the analyzes was controlled using a certified reference material (ERM®-BB422, JRC Institute for Reference Materials and Measurements, Geel, Belgium) – fish muscle and BCR-186 (IRMM, Geel, Belgium), lyophilised pig kidney. The concentrations determined in the reference material were within the permissible deviations specified in the obtained certificate and are shown in Table 3.

**Table 3.** Certified concentration and measured values for the Certified Reference Material ERM®-BB422 (fish muscle) and BCR-186 (lyophilized pig kidney) (mg/kg)

Element	Assigned values (mg/kg)	Measured value ± Uncertainty (mg/kg)
Pb	0.306±0.011	0.298±0.089
Cd	0.0075±0.0018	0.0070±0.0019
Hg	0.601±0.030	0.615±0.150

The concentrations in the tested samples were compared with the maximum allowed concentrations (MAC) in fish meat established by the Regulations. According to the

legislation of the European Union [33] and the national legislation [34], the maximum allowed concentrations for Pb, Cd and Hg in fresh fish meat are 0.30, 0.05 and 0.50 mg/kg, respectively.

## Statistical Analyses

Statistical analysis of the results was elaborated using Microsoft Office Excel 2010 and GraphPad Prism software, version 9.00 for Windows (GraphPad Software, San Diego, California USA, [www.graphpad.com](http://www.graphpad.com)). All parameters were described by mean and standard deviation (SD). One-way ANOVA with Tukey's post hoc test was performed to assess the significance of differences between heavy metal concentrations during a two-year period in spring and autumn. To determine the significance of the differences between heavy metal concentrations at two locations, t-test was used. Differences were considered significant if  $p < 0.01$  or  $p < 0.05$  were observed.

## RESULTS

The total concentration of heavy metals (mg/kg) in the muscle tissue of certain fish species in the territories before Belgrade (sampling point A) and in Belgrade (sampling point B) during the two-year period of investigation is shown in Table 4.

**Table 4.** Total concentrations of heavy metals (mg/kg) in the muscle tissue of selected fish species from the Sava River, aggregated across sampling points A and B (n=12) during the two-year study period

Type of fish	Parameter	$\bar{X} \pm Sd$			
		Autumn '23	Spring '24	Autumn '24	Spring '25
Catfish	Pb	0.215 <sup>A,B</sup> ±0.042	0.282 <sup>ACD</sup> ±0.068	0.113 <sup>BCE</sup> ±0.0163	0.187 <sup>DE</sup> ±0.038
	Cd	0.069 <sup>A</sup> ±0.020	0.087 <sup>B</sup> ±0.025	0.082 <sup>C</sup> ±0.012	0.032 <sup>ABC</sup> ±0.015
	Hg	0.278±0.106	0.303±0.116	0.261±0.088	0.289±0.088
Carp	Pb	0.212 <sup>A</sup> ±0.045	0.248 <sup>B</sup> ±0.056	0.115 <sup>ABC</sup> ±0.025	0.245 <sup>C</sup> ±0.062
	Cd	0.086 <sup>A</sup> ±0.020	0.103 <sup>BC</sup> ±0.030	0.077 <sup>BD</sup> ±0.015	0.021 <sup>ACD</sup> ±0.008
	Hg	0.212±0.106	0.243 <sup>a</sup> ±0.126	0.268 <sup>A</sup> ±0.098	0.133 <sup>Aa</sup> ±0.038
Perch	Pb	0.111 <sup>A</sup> ±0.026	0.088 <sup>B</sup> ±0.025	0.090 <sup>C</sup> ±0.038	0.225 <sup>ABC</sup> ±0.083
	Cd	0.033±0.010	0.038±0.012	0.041±0.015	0.041±0.014
	Hg	0.154±0.061	0.201 <sup>A</sup> ±0.079	0.201 <sup>B</sup> ±0.075	0.090 <sup>AB</sup> ±0.038
Prussian carp	Pb	0.134±0.035	0.148±0.051	0.164±0.036	0.177±0.050
	Cd	0.047±0.007	0.041±0.013	0.036±0.008	0.045±0.022
	Hg	0.112 <sup>AB</sup> ±0.026	0.125 <sup>a</sup> ±0.034	0.161 <sup>A</sup> ±0.037	0.163 <sup>Ba</sup> ±0.037

**Legend:** Same letter in row <sup>a</sup> –  $p < 0.05$ ; <sup>ABCDE</sup> –  $p < 0.01$ .

A significantly higher ( $p < 0.01$ ) concentration of lead in the muscle tissue of catfish was found in the spring of 2024 ( $0.282 \pm 0.068$  g/kg) compared to the fall of 2023 ( $0.215 \pm 0.042$  mg/kg), as well as in the spring of 2025 ( $0.187 \pm 0.038$  mg/kg) and the fall of 2024 ( $0.113 \pm 0.0163$  mg/kg). Also, the concentration of lead in the muscle tissue of catfish in the spring of 2025 was significantly higher ( $p < 0.01$ ) than in the fall of 2024. In the fall of 2023, the concentration of lead in the examined catfish tissue was significantly higher ( $p < 0.01$ ) compared to the fall of the following year. In the spring of 2025, a significantly lower ( $p < 0.01$ ) cadmium concentration was found in the muscle tissue of catfish ( $0.032 \pm 0.015$  mg/kg) compared to other comparison periods. Between the other periods of comparison, no significant differences were found in the concentration of cadmium in the examined catfish tissue. The concentration of mercury in the muscle tissue of catfish was generally uniform during the two-year study and no statistically significant differences were found between the compared periods.

The concentration of lead in the muscle tissue of carp in the fall of 2024 ( $0.115 \pm 0.025$  mg/kg) was significantly lower ( $p < 0.01$ ) compared to the concentration of this heavy metal in all other comparison periods, while no significant differences were found between the compared concentrations in the other periods during the two-year study. In the spring of 2025, a significantly lower ( $p < 0.01$ ) concentration of cadmium ( $0.021 \pm 0.008$  mg/kg) was found compared to the previous period (autumn 2023-autumn 2024), while in the spring of the previous year, the concentration of cadmium in the muscle tissue of carp ( $0.103 \pm 0.030$  mg/kg) was significantly higher ( $p < 0.01$ ) compared to the concentration of cadmium in the examined carp tissue in the fall of 2024 ( $0.077 \pm 0.015$  mg/kg). Similar results were obtained for mercury concentration in carp muscle tissue. In the spring of 2025, a significantly lower concentration of this heavy metal was found in the muscle tissue of carp ( $0.133 \pm 0.038$  mg/kg) compared to the spring of 2024 ( $0.243 \pm 0.126$  mg/kg) ( $p < 0.05$ ) and the fall of 2024 ( $0.268 \pm 0.098$  mg/kg) ( $p < 0.01$ ).

In the muscle tissue of perch, the lead concentration was significantly higher ( $p < 0.01$ ) in the spring of 2025 ( $0.225 \pm 0.083$  mg/kg) compared to the other three test periods. During the two-year study period, no significant differences were found between the concentration of cadmium in the muscle tissue of perch in all four comparison periods. The concentration of mercury in the muscle tissue of perch ( $0.090 \pm 0.038$  mg/kg) was significantly lower ( $p < 0.01$ ) in the spring of 2025 compared to the spring and autumn of 2024 ( $0.201 \pm 0.079$  mg/kg;  $0.201 \pm 0.075$  mg/kg, respectively).

The concentration of lead and cadmium in the muscle tissue of Prussian carp did not change significantly during the two-year test period, while the concentration of mercury in the examined tissue of Prussian carp was significantly lower ( $p < 0.01$ ) in the fall of 2023 ( $0.112 \pm 0.026$  mg/kg) compared to the mercury concentration in the fall of the following year ( $0.161 \pm 0.037$  mg/kg) and the spring of 2025 ( $0.163 \pm 0.037$  mg/kg). Also, significant differences ( $p < 0.05$ ) were found in the concentration of this heavy metal in the muscle tissue of Prussian carp in the spring of 2024 ( $0.125 \pm 0.034$



mg/kg) and the spring of 2025 ( $0.163 \pm 0.037$  mg/kg), where the concentration increased significantly during one year.

Table 5 shows the concentrations of lead, cadmium and mercury in the muscle tissue of catfish (mg/kg) at the sampling point A and sampling point B during a two-year period. In the muscle tissue of catfish, no significant differences were found between the concentration of lead in the territory of the catch upstream of Belgrade and in Belgrade during the entire period of investigation. In the spring of 2025 at the sampling point B, a significantly higher ( $p < 0.05$ ) concentration of cadmium ( $0.041 \pm 0.015$  mg/kg) was found in the muscle tissue of catfish than in muscle tissue of catfish caught at the sampling point A ( $0.024 \pm 0.009$  mg/kg). In the other periods, no significant differences were found between the concentrations of cadmium in the muscle tissue of catfish in the two compared catch territories. The mercury concentration in the muscle tissue of catfish caught in sampling point B was significantly higher ( $p < 0.01$ ) in all four comparison periods ( $0.355 \pm 0.091$  mg/kg;  $0.385 \pm 0.092$  mg/kg;  $0.331 \pm 0.065$  mg/kg;  $0.364 \pm 0.053$  mg/kg, respectively) compared to the mercury concentration in the same catfish tissue caught at the sampling point A ( $0.201 \pm 0.049$  mg/kg;  $0.222 \pm 0.073$  mg/kg;  $0.192 \pm 0.035$  mg/kg;  $0.213 \pm 0.023$  mg/kg, respectively).

**Table 5.** Concentration of examined heavy metals (Pb, Cd, Hg) in muscle tissue of catfish (mg/kg) from the Sava River at sampling point A and sampling point B (n=6) during the two-year study period

Parameter	Location	$\bar{X} \pm Sd$			
		Autumn '23	Spring '24	Autumn '24	Spring '25
Pb	Sampling point A	$0.204 \pm 0.039$	$0.270 \pm 0.076$	$0.110 \pm 0.014$	$0.178 \pm 0.037$
	Sampling point B	$0.227 \pm 0.047$	$0.294 \pm 0.064$	$0.118 \pm 0.019$	$0.196 \pm 0.040$
Cd	Sampling point A	$0.070 \pm 0.024$	$0.096 \pm 0.029$	$0.086 \pm 0.014$	$0.024^a \pm 0.009$
	Sampling point B	$0.069 \pm 0.017$	$0.077 \pm 0.019$	$0.079 \pm 0.009$	$0.041^a \pm 0.015$
Hg	Sampling point A	$0.201^A \pm 0.049$	$0.222^A \pm 0.073$	$0.192^A \pm 0.035$	$0.213^A \pm 0.023$
	Sampling point B	$0.355^A \pm 0.091$	$0.385^A \pm 0.092$	$0.331^A \pm 0.065$	$0.364^A \pm 0.053$

**Legend:** Same letter in row <sup>a</sup> –  $p < 0.05$ ; <sup>A</sup> –  $p < 0.01$ .



During the two-year study period, the concentrations of lead, cadmium and mercury in the muscle tissue of carp (mg/kg) caught in the sampling point A and sampling point B were compared (Table 6). The concentration of lead in the examined tissue of carp caught in the sampling point A was not significantly different from the concentration of lead in the same tissue of this species of fish caught in the sampling point B during the four comparison periods. A similar situation was found when comparing the concentration of cadmium in the muscle tissue of carp, where during the two-year period of testing, no significant differences were found between the concentrations of the examined heavy metal in carp caught in the sampling point A and sampling point B, except for the spring of 2025 when a significantly higher  $p<0.01$  value of cadmium was found at the sampling point B. The concentration of mercury was significantly higher ( $p<0.01$ ) in the muscle tissue of the examined species of fish at sampling point B in the fall of 2023 ( $0.303\pm0.068$  mg/kg), spring of 2024 ( $0.343\pm0.094$  mg/kg) and autumn of the same year ( $0.355\pm0.049$  mg/kg) in relation to the concentration of this heavy metal in the muscle tissue of carp caught at sampling point A during the compared test periods ( $0.122^A\pm0.025$  mg/kg;  $0.143^A\pm0.046$  mg/kg;  $0.180^A\pm0.019$  mg/kg, respectively). In the spring of 2025, no significant differences were found between the concentration of mercury in the muscle tissue of the investigated species of fish spawned at the sampling points A and B.

**Table 6.** Concentration of examined heavy metals (Pb, Cd, Hg) in muscle tissue of carp (mg/kg) from the Sava River at sampling point A and sampling point B (n=6) during the two-year study period

Parameter	Location	$\bar{X} \pm Sd$			
		Autumn '23	Spring '24	Autumn '24	Spring '25
Pb	Sampling point A	0.200±0.047	0.232±0.056	0.104±0.022	0.212±0.049
	Sampling point B	0.224±0.043	0.263±0.057	0.127±0.024	0.278±0.060
Cd	Sampling point A	0.081±0.020	0.097±0.029	0.072±0.015	0.016 <sup>A</sup> ±0.005
	Sampling point B	0.092±0.019	0.110±0.032	0.081±0.015	0.027 <sup>A</sup> ±0.006
Hg	Sampling point A	0.122 <sup>A</sup> ±0.025	0.143 <sup>A</sup> ±0.046	0.180 <sup>A</sup> ±0.019	0.123±0.037
	Sampling point B	0.303 <sup>A</sup> ±0.068	0.343 <sup>A</sup> ±0.094	0.355 <sup>A</sup> ±0.049	0.144±0.040

**Legend:** Same letter in row <sup>A</sup> –  $p<0.01$ .

The concentrations of lead, cadmium and mercury in the muscle tissue of perch (mg/kg) caught at the sampling points A and B during a two-year period are shown in Table 7. No significant differences were found in the concentration of lead and cadmium in the muscle tissue of this type of fish caught in the observed locations, except for the last two examined periods when a significant difference was determined for lead. However, the concentration of mercury in the muscle tissue of perch caught at the sampling point A was significantly higher ( $p<0.01$ ) in the fall of 2023 ( $0.201\pm0.044$  mg/kg), spring ( $0.262\pm0.057$  mg/kg) and fall of 2024 ( $0.259\pm0.063$  mg/kg) in relation to the concentration of mercury in the muscle tissue of the examined species of fish caught at the sampling point B in the compared periods of the study ( $0.107\pm0.031$  mg/kg;  $0.140\pm0.040$  mg/kg;  $0.144\pm0.022$  mg/kg, respectively). Completely opposite results were obtained in the spring of 2025, where the mercury concentration in the examined tissue of perch caught in the sampling point A ( $0.056\pm0.007$  mg/kg) was significantly lower ( $p<0.01$ ) compared to the mercury concentration of perch caught at sampling point B ( $0.124\pm0.018$  mg/kg).

**Table 7.** Concentration of examined heavy metals (Pb, Cd, Hg) in muscle tissue of perch (mg/kg) from the Sava River at sampling point A and sampling point B (n=6) during the two-year study period

Parameter	Location	$\bar{X} \pm Sd$			
		Autumn '23	Spring '24	Autumn '24	Spring '25
Pb	Sampling point A	$0.099\pm0.022$	$0.078\pm0.022$	$0.056^A\pm0.007$	$0.292^A\pm0.053$
	Sampling point B	$0.123\pm0.026$	$0.099\pm0.024$	$0.124^A\pm0.018$	$0.157^A\pm0.036$
Cd	Sampling point A	$0.028\pm0.009$	$0.032\pm0.008$	$0.037\pm0.016$	$0.037\pm0.016$
	Sampling point B	$0.037\pm0.008$	$0.044\pm0.014$	$0.046\pm0.013$	$0.046\pm0.013$
Hg	Sampling point A	$0.201^A\pm0.044$	$0.262^A\pm0.057$	$0.259^A\pm0.063$	$0.056^A\pm0.007$
	Sampling point B	$0.107^A\pm0.031$	$0.140^A\pm0.040$	$0.144^A\pm0.022$	$0.124^A\pm0.018$

**Legend:** Same letter in row <sup>A</sup> –  $p<0.01$ .

In the spring of 2024, the concentration of lead in the muscle tissue of Prussian carp caught at the sampling point A ( $0.177\pm0.053$  mg/kg) was statistically significantly higher ( $p<0.05$ ) compared to the concentration of lead in the same tissue of Prussian carp caught in the sampling point B ( $0.120\pm0.031$  mg/kg). The concentration of this heavy metal in the examined tissue of Prussian carp caught at the observed locations

was not significantly different in the other three periods of comparison (autumn 2023, autumn 2024 and spring 2025) (Table 8). No significant differences were found between the concentration of cadmium in the muscle tissue of this species of fish caught in the sampling points A and B. In the muscle tissue of Prussian carp caught at the sampling point B, the concentration of mercury ( $0.131\pm0.019$  mg/kg;  $0.144\pm0.031$  mg/kg;  $0.191\pm0.019$  mg/kg;  $0.194\pm0.020$  mg/kg, respectively) was significantly higher ( $p<0.05$ ;  $p<0.01$ ) compared to the muscle tissue of Prussian carp caught at sampling point A ( $0.093\pm0.016$  mg/kg;  $0.107\pm0.027$  mg/kg;  $0.131\pm0.021$  mg/kg;  $0.132\pm0.018$  mg/kg, respectively) during all four comparison periods.

**Table 8.** Concentration of examined heavy metals (Pb, Cd, Hg) in muscle tissue of Prussian carp (mg/kg) from the Sava River at sampling point A and sampling point B (n=6) during the two-year study period

Parameter	Location	$\bar{X} \pm Sd$			
		Autumn '23	Spring '24	Autumn '24	Spring '25
Pb	Sampling point A	$0.149\pm0.041$	$0.177^a\pm0.053$	$0.181\pm0.041$	$0.200\pm0.061$
	Sampling point B	$0.119\pm0.021$	$0.120^a\pm0.031$	$0.147\pm0.021$	$0.154\pm0.020$
Cd	Sampling point A	$0.043\pm0.004$	$0.038\pm0.012$	$0.034\pm0.008$	$0.037\pm0.015$
	Sampling point B	$0.051\pm0.008$	$0.045\pm0.014$	$0.039\pm0.008$	$0.053\pm0.026$
Hg	Sampling point A	$0.093^A\pm0.016$	$0.107^a\pm0.027$	$0.131^A\pm0.021$	$0.132^A\pm0.018$
	Sampling point B	$0.131^A\pm0.019$	$0.144^a\pm0.031$	$0.191^A\pm0.019$	$0.194^A\pm0.020$

**Legend:** Same letter in row <sup>a</sup> –  $p<0.05$ ; <sup>A</sup> –  $p<0.01$ .

DISCUSSION

Investigation of the presence of heavy metals in the muscle tissue of four species of freshwater fish (catfish – *Silurus glanis*, carp – *Cyprinus carpio*, perch – *Sander lucioperca*, Prussian carp – *Carassius auratus gibelio*) from the territory of the Sava River at sampling points A and B, indicated certain spatial and seasonal differences, especially in relation to the concentration of mercury and cadmium. The overall results indicate a moderate, but clearly detected influence of the urban area on the quality of the river ichthyofauna, considering the different response of the observed fish species regarding the accumulation of lead, cadmium and mercury.

Lead was present in most samples in concentrations that did not exceed legally permitted values and did not show significant differences between locations, except for Prussian carp in spring 2024, where the lead level was higher at sampling point A, as well as perch in autumn 2024 and spring 2025, where higher and lower values were observed at the sampling point B, respectively. This result may indicate localized sources of contamination, including soil erosion and agricultural sources, which is consistent with research in the region [35]. Also, seasonal fluctuations of lead concentrations were observed in catfish and perch, which may indicate changes in the biological availability of the metal due to hydrological and temperature factors. Perch, especially in the spring of 2025, showed a significantly higher concentration of lead, which may be related to the increased mobilization of the metal from the sediments due to the rise in the water level in the spring months.

Cadmium was detected in the majority of samples in low concentrations, but unlike lead, exceeding the maximum allowed concentration (0.05 mg/kg) was recorded in catfish, carp and Prussian carp in several seasons. The values of cadmium in carp were above the MAC in autumn 2023 (0.086 mg/kg), spring 2024 (0.103 mg/kg) and autumn 2024 (0.077 mg/kg), while in catfish, exceedances were recorded in autumn 2023 (0.069 mg/kg), spring 2024 (0.087 mg/kg) and autumn in 2024 (0.082 mg/kg). In the case of Prussian carp, exceedances were determined in the fall of 2023 (0.047 mg/kg close to the limit value), but they were especially pronounced in sampling point B in the fall of 2023 (0.051 mg/kg) and spring of 2025 (0.053 mg/kg). This makes cadmium stand out as the riskiest metal in this research, which indicates a potential health risk for consumers and the need for strict monitoring. Although in the spring of 2025 the concentration of cadmium was significantly lower in catfish and carp (0.032 and 0.021 mg/kg), the previously determined exceedances cannot be ignored. Similar findings of cadmium accumulation in fish downstream of urban and industrial zones have been described in other papers [36,37]. It is worth noting that cadmium is deposited to a greater extent in the liver and kidneys, and the analysis of those tissues could give an even more precise insight into the actual degree of exposure.

Unlike the concentration of lead and cadmium, the most pronounced spatial differences were found in the concentration of mercury. In catfish, carp and Prussian carp, the concentration of mercury was significantly higher in samples from sampling point B during several seasons. This indicates a clear anthropogenic influence of urban and industrial sources. Given that methylmercury is particularly toxic and has a high potential for bioaccumulation and biomagnification in the trophic chain, these results are particularly significant from the point of view of consumer protection and ecosystem preservation. Similar findings were reported in large rivers in Europe and Asia, where Hg concentrations were significantly higher in benthic and omnivorous species [38,39]. The increased concentration of mercury in samples downstream from urban areas was also confirmed in works indicating that urban wastewater and atmospheric deposition are key sources of this metal in aquatic ecosystems [37,40]. It is interesting that in the case of perch, as a higher predator, the opposite trend was

recorded – higher concentrations of mercury were present on the sampling point A during three of the four analyzed periods. This phenomenon may be the result of localized sources of pollution or changes in prey availability, as perch show high sensitivity to disturbances in the food chain. It is also possible that perch in this part of the river feed on species that migrate from tributaries with greater contamination, which further complicates the interpretation of spatial patterns. In addition, a significant seasonal oscillation of mercury concentration was observed in perch, which may be related to its migration patterns, changes in diet or metabolism depending on temperature and reproductive cycle [41].

Based on the obtained results, it can be concluded that the concentrations of lead and mercury in the largest number of samples were in accordance with the prescribed limit values, while cadmium repeatedly exceeded the maximum allowed concentration in the muscle tissue of carp, catfish and Prussian carp. This indicates that the fish from this part of the river Sava cannot be considered completely safe for human consumption, especially when it comes to species that show a tendency to accumulate cadmium. Therefore, it is necessary to introduce continuous monitoring, expand and confirm toxicological analyzes on a larger number of fish samples, including the monitoring of other tissues and biomarkers of exposure. The obtained results supplement and confirm findings from earlier research in Serbia and the region [4,5] and indicate the importance of a multispecies approach in assessing the state of aquatic ecosystems, because different fish species react differently to the same stressors in the environment. These findings further reinforce the need for an integrated analysis that includes physicochemical, biological, and toxicological parameters for a complete assessment of the ecosystem health of river courses in urban areas.

## **CONCLUSION**

The research results show that the presence of heavy metals in freshwater fish from the Sava River reflects the influence of various anthropogenic and natural factors. While lead and mercury concentrations were generally within legal limits, cadmium exceeded permissible values in catfish, carp, and Prussian carp, indicating a potential health risk. Variations between species and seasons suggest complex bioaccumulation patterns, highlighting the need for systematic monitoring of heavy metals in fish to ensure food safety and protect aquatic ecosystems.

## **Acknowledgements**

The study was supported by the Ministry of Science, Technological Development and Innovation of the Republic of Serbia (Contract number 451-03-136/2025-03/200143).


## Authors' contributions

DJ and JJ participated in conducting the experiment and drafting the manuscript. RM participated in the design of the experiment. DP, SR and VD participated in conducting the experiment. JJ performed the statistical analysis of the experiment results. SG participated as coordinator in all activities. DŠ designed the study, participated in drafting the manuscript and was coordinator of all activities. All authors read and approved the final manuscript.


## Declaration of competing interests


The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.


## ORCID iDs


Radmila Marković  <https://orcid.org/0000-0001-8467-0551>


Svetlana Grdović  <https://orcid.org/0000-0003-4445-2247>

Jelena Janjić  <https://orcid.org/0000-0003-3351-7199>

Dragoljub Jovanović  <https://orcid.org/0000-0003-1665-7106>

Dejan Perić  <https://orcid.org/0000-0002-4752-7489>

Vladimir Dimitrijević  <https://orcid.org/0000-0002-3593-7130>

Stamen Radulović  <https://orcid.org/0000-0002-7250-537X>

Dragan Šefer  <https://orcid.org/0000-0002-4394-6336>

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## MONITORING SADRŽAJA TEŠKIH METALA U MIŠIĆNOM TKIVU RIBA DUŽ REKE SAVE U SRBIJI

Radmila MARKOVIĆ, Svetlana GRDOVIĆ, Jelena JANJIĆ,  
Dragoljub JOVANOVIĆ, Dejan PERIĆ, Vladimir DIMITRIJEVIĆ,  
Stamen RADULOVIĆ, Dragan ŠEFER

Cilj ove studije bio je ispitivanje prisustva teških metala (Pb, Cd i Hg) u mišićnom tkivu ribe iz reke Save, sa dve lokacije: pre Beograda u blizini Obrenovca i u Beogradu. Radi utvrđivanja sadržaja teških metala u mesu riba, sakupljeno je 196 uzoraka različitih vrsta riba: babuške, šarana, smuđa i soma, tokom jeseni i proleća 2023. – 2025. god. Za određivanje sadržaja teških metala korišćena je mikrotalasna peć i atomski apsorpcioni spektrometar. Najveći prosečan sadržaj Pb ( $0.282 \pm 0.068$  mg/kg) i Hg ( $0.303 \pm 0.116$  mg/kg) pronađen je u mišićnom tkivu soma, dok je najmanji prosečan sadržaj Pb i Hg utvrđen u mišićnom tkivu smuđa ( $0.088 \pm 0.025$  mg/kg), ( $0.090 \pm 0.038$  mg/kg), redom. Najveći prosečan sadržaj Cd zabeležen je u mišićnom tkivu šarana ( $0.103 \pm 0.030$  mg/kg), dok je najmanji utvrđen u mišićnom tkivu soma ( $0.032 \pm 0.015$  mg/kg). U svim ispitivanim uzorcima utvrđene koncentracije Pb i Hg su bile ispod maksimalno dozvoljenih koncentracija koje su regulisane Pravilnikom Republike Srbije i propisima Evropske Unije. Utvrđene koncentracije kadmijuma u mišićnom tkivu šarana, soma i babuške su u više navrata bile iznad maksimalno dozvoljenih koncentracija od 0.05 mg/kg sveže ribe. Riba iz ovog dela Save ne može se u potpunosti smatrati bezbednom za ljudsku ishranu, posebno kada je reč o vrstama koje pokazuju sklonost ka akumulaciji Cd. Uopšteno, sezonske varijacije su bile očiglednije od prostornih razlika, pri čemu je nekoliko vrsta pokazalo umereno veće koncentracije metala u proleće, posebno 2025. godine, što ukazuje na suptilne, ali ponavljajuće sezonske tendencije. Razlike između teritorija pre Beograda i unutar Beograda bile su generalno male, iako su koncentracije žive bile konstantno više na nizvodnoj lokaciji, što ukazuje na blag, ali stabilan prostorni uticaj.