

SELENIUM STATUS OF FEEDSTUFFS AND GRAZING EWES IN SERBIA

VALČIĆ OLIVERA, JOVANOVIĆ I, MILANOVIĆ SVETLANA and GVOZDIĆ D

University of Belgrade, Faculty of Veterinary Medicine, Serbia

(Received 1st March 2013)

A total of 221 feedstuff samples (187 grain samples and 34 hay samples) were collected from 13 different locations on the territory of the Republic of Serbia. The results expressed as $\mu\text{g}/\text{kg}$ have shown an average selenium content in grain samples of 34.3 ± 17.1 and 53.8 ± 18.7 in hay samples. However, a distinct difference was noted between samples collected from the locations north of the river Danube (52.8 ± 20.0 and 73.4 ± 21.3 for grain and hay samples, respectively) compared to the locations south of the Danube (23.3 ± 15.4 and 41.7 ± 17.1 for grain and hay samples, respectively).

GPx activity was measured in a total of 58 blood plasma samples collected from ewes grazing on 5 different locations. The average GPx activity was $157.4 \pm 61.9 \mu\text{kat}/\text{L}$ and it followed a pattern similar to the distribution of Se in feedstuffs. GPx activity was higher in samples collected north of the Danube ($212.8 \pm 91.2 \mu\text{kat}/\text{L}$), compared to ewes south of the Danube ($66.9 \pm 14.0 \mu\text{kat}/\text{L}$).

According to the obtained results and literature data Serbia can be described as a selenium deficient area, showing a marked deficiency on the locations south of the Danube.

Key words: feedstuffs, glutathione peroxidase, selenium, Serbia, sheep

INTRODUCTION

Since in 1817 Berzelius discovered selenium (Se) it has been considered only for its highly toxic properties, until Schwarz and Foltz (1958) showed that in extremely small quantities it performed as an essential nutrient. Schwarz's study stimulated an astonishing amount of research on the impact of selenium in plants and animals physiology and health.

Soils and rocks are the ultimate source of all selenium in the food chain of plants and land animals. Most soils contain from 0.1 to 2 mgSe/kg (Swaine, 1955), but these values can go up to 100 mgSe/kg. It is important to be able to recognize and distinguish areas of selenium deficiency, as well as toxicity.

The estimation of selenium in localized areas can be approached by soil chemical analysis and may be used as an indicator of selenium levels in locally grown plants, or animals consuming them, but such an approach has serious drawbacks. The biological availability of selenium to plants, and subsequently to animals, depends on a number of factors such as the chemical form in which selenium is present, soil pH, mechanical soil properties (Gissel–Nielsen, 1971), and use of mineral fertilizers (Gissel–Nielsen, 1974). After absorption of selenite or selenate ions, plants synthesize selenoamino acids with selenomethionine (SeMet) representing more than 50% of the Se present in cereal grains (Olson and Palmer, 1976) with Se-methyl–selenomethionine, selenocysteine, Se-methyl-selenocysteine being the other selenocompounds found in plants (Brody, 1994). Plant species are another consideration as some types of plants are more efficient at absorbing selenium from the soil than others (Rosenfield and Beath, 1964). Most forage, cereal and oilmeal crop plants are non-accumulator plants containing on average less than 25 mgSe/kg, dry weight do not accumulate over 100 mg/kg even when grown on seleniferous soils (Terry *et al.*, 2000)

The need for selenium geobotanical mapping arose soon after Kubota *et al.* (1967) produced a detailed map of the USA which clearly defined areas where selenium toxicity or deficiency may be expected. An extensive study on more than 11,000 feedstuff samples on 1,103 locations in 28 provinces in the People's Republic of China revealed serious selenium deficiency as 25% of the tested samples contained less than 20 µgSe/kg (Liu *et al.*, 1986). Geobotanical mapping of Europe has revealed areas which are deficient in selenium (Scandinavian Peninsula), as well as sporadic zones of selenium toxicity (Ireland). The situation on the Balkan Peninsula is far from ideal. Studies carried out during the 90-ties revealed low selenium concentrations in cereal samples from Macedonia (Mihailović *et al.*, 1993) and extremely low (all forage samples were below 50 µgSe/kg) in the region of south Serbia on the Pester highland.

Grazing animals, such as sheep and goats, which in extensive production settings rarely are supplemented with professionally controlled amounts of micronutrients, are the first to reflect low selenium conditions. Selenium nutrition of ruminants has some specific features, which create specific problems. Indeed, rumen bacteria incorporate selenium during protein synthesis; hence it is absorbed as selenoaminoacids.

Selenium functions within mammalian systems primarily in the form of selenoproteins. Selenoproteins contain selenium as selenocysteine and perform a variety of physiological roles. The most important identified selenoproteins include: 5 isoforms of glutathione peroxidase (GPx) selenoprotein P; iodothyronine deiodinase types 1, 2, and 3; selenoprotein W; thioredoxin reductases; and selenophosphate synthetase. The principal methods for the assessment of selenium status include the determination of its concentration and measurement of GPx activities in tissues, as well as the occurrence of selenium responsive diseases in animals. The activity of erythrocyte GPx reflects the long-standing

selenium status of the animal, as the synthesis of the enzyme occurs only during the time of erythropoiesis due to the fact that mature erythrocytes do not have the organelles needed for protein synthesis (Osame *et al.*, 1990). Thus, whole blood GPx activity can be considered as the reflection of long-time selenium status and plasma/serum GPx activity reflects short-term changes as the distinct blood compartments reflect different time frames in the nutritional history of the animal (Waldner *et al.*, 1998). This was moreover confirmed by van Ryssen *et al.* (2013) who studied the time-dependent effect of selenium supplementation on the relationship between selenium concentrations in whole blood and plasma of sheep. The authors observed that the time needed to achieve a stabilized ratio between plasma and whole blood selenium was on average 50 days, as whole blood needed a long time span in order to incorporate selenium into the erythrocytes. Wilson and Judson (1976) reported that GPx activity in the erythrocytes is more resistant to the effect of storage time than plasma GPx. Besides, Sheperd and Miller (1981) disclosed that sheep erythrocyte GPx activity is 99 times higher than in the plasma, thus confirming the previous report by Scholtz and Hutchinson (1979) who recorded that out of the total GPx activity only 0.70% is due to plasma GPx.

Glutathione peroxidase plays a major role in the protection of cells against oxidative damage by endogenous peroxides. Peroxides can damage cell membranes leading to degeneration and necrosis.

Selenium deficiency in domestic animals manifests itself in a number of disorders which result in poor health, and low productive and reproductive performances. In sheep, two selenium-responsive conditions are recognized: one is a myopathy which presents itself in lambs as white muscle disease (WMD) (Muth *et al.*, 1958), the other is a syndrome of lowered productivity which ranges from poor wool production to clinically manifested selenium responsive unthriftiness (Drake *et al.*, 1960).

The aim of this study was to determine selenium content in forage and cereals grown in Serbia in order to develop a geobotanical map of the region. Grazing, selenium non supplemented sheep were selected for the determination of selenium dependent GPx activity in order to detect areas of possible selenium deficiency in several agriculturally important regions.

MATERIAL AND METHODS

Feedstuff samples

Samples of feedstuffs (187 of grain and 34 of hay) were collected from 13 locations throughout Serbia, on farms belonging to local farmers. The fields were never previously treated with Se-containing fertilizers. All grain samples (wheat, corn, barley, and oats) were cleaned from foliage, stalks, roots, soil, grit and non edible parts. Hay was sampled from the central portion of the hay stack. Samples

were stored in paper-bags at room temperature until analysis. Selenium content was determined by the fluorometric method (Lindberg, 1968)

Experimental animals

The study was conducted on 5 locations in Serbia, on a total of 58 ewes. Groups of selected animals (Wirtenberg x Cigaja crossbred sheep), were formed on each location on the basis of a common age range (3-5 years) and sex (female), multiparous, and feeding on locally grown forage and grain.

After puncture of *v. jugularis* blood samples were collected in heparinized test tubes and stored at 8°C for not more than 12h. Immediately before the measurement of GPx activity 50 µL of full blood samples were hemolysed in 1mL Drabkin's reagent, thus making a 21x dilution of hemolysate. Selenium dependent GPx activity was determined within 24h from sampling. Glutathione peroxidase (GPx – EC. 1.11.1.9) activity was analyzed spectrophotometrically by the coupled test (Günzler *et al.*, 1974) using a concentration of tertiary butyl hydroperoxide (TBH) below 2.32 mM in order to measure only the activity of selenium dependent GPx (Sankari and Atroshi, 1983).

RESULTS

Selenium concentrations in feedstuffs sampled on a total of 13 locations in Serbia are shown in Table 1. The average selenium content in grain and hay samples in Serbia was 34.3 ± 17.1 µgSe/kg, and 53.8 ± 18.8 µgSe/kg, respectively. The highest concentration of selenium (52.8 ± 20.0 µgSe/kg) in grains was determined in the samples collected on locations north of the river Danube. In this region the highest value was recorded in Melenci (60.0 ± 21.0 µgSe/kg), and lowest in Vršac (44.1 ± 33.0 µgSe/kg). Selenium content in hay samples showed marked differences and ranged from 104.6 ± 20.0 µgSe/kg on location Bačka Topla to 46.6 ± 15.9 µgSe/kg in Apatin. The average selenium content in hay samples collected north of the river Danube (73.4 ± 21.3 µgSe/kg) was higher compared to the locations south of the Danube (41.7 ± 17.2 µgSe/kg).

In Central Serbia the concentration of selenium in feedstuffs was markedly lower (23.3 ± 15.4 µgSe/kg), ranging from 4.3 ± 1.0 µgSe/kg in grain samples on the Sjenica location to 26.0 ± 21.4 µgSe/kg on location Šabac.

The average GPx activity in ewes whole blood sampled on five locations in Serbia was 157.4 ± 61.9 µkat/L. GPx activity was higher in the samples taken from ewes grazing north of the Danube compared from samples on locations south of the Danube (Table 2.) The lowest recorded activities were in whole blood samples from the location Aleksinac (65.4 ± 11.1 µkat/L).

Table 1. Selenium content ($\mu\text{g}/\text{kg}$) in grain and hay feedstuff samples in Serbia

Location	Grain ($\mu\text{g}/\text{kg}$)		Hay ($\mu\text{g}/\text{kg}$)	
North of Danube				
Subotica	58.3 \pm 18.6	(n=11)	-	
Bačka Topola	47.2 \pm 17.0	(n=25)	104.6 \pm 20.0	(n=5)
Apatin	51.2 \pm 27.1	(n=11)	46.6 \pm 15.9	(n=5)
Melenci	60.0 \pm 21.0	(n=4)	-	
Zrenjanin	59.0 \pm 13.0	(n=4)	-	
Vršac	44.0 \pm 33.0	(n=4)	66.2 \pm 31.6	(n=3)
Futog	58.9 \pm 18.5	(n=11)	-	
North	52.8 \pm 20.0	(n=70)	73.4 \pm 21.3	(n=13)
South of Danube				
Ruma	36.8 \pm 16.7	(n=17)	66.4 \pm 30.3	(n=3)
Šabac	26.0 \pm 21.4	(n=24)	-	
Smederevo	18.9 \pm 13.4	(n=54)	45.9 \pm 21.6	(n=8)
Zajačar	25.8 \pm 18.7	(n=9)	42.0 \pm 13.3	(n=6)
Kraljevo	24.9 \pm 12.9	(n=8)	-	
Sjenica	4.3 \pm 1.0	(n=5)	14.4 \pm 4.0	(n=4)
South	23.3 \pm 15.4	(n=117)	41.7 \pm 17.1	(n=21)
SERBIA	34.3 \pm 17.1	(n=187)	53.8 \pm 18.7	(n=34)

Table 2. Whole blood glutathione peroxidase activity ($\mu\text{kat}/\text{L}$) in ewes in Serbia, according to location

Location	GPx ($\mu\text{kat}/\text{L}$)	
North of Danube		
Kač		
Zrenjanin	236.5 \pm 85.7	(n=15)
Čoka	194.0 \pm 101.1	(n=14)
North	212.8 \pm 91.2	(n=36)
South of Danube		
Ub	70.2 \pm 20.2	(n=7)
Aleksinac	65.4 \pm 11.1	(n=15)
South	66.9 \pm 14.0	(n=22)
SERBIA	157.4 \pm 61.9	(n=58)

DISCUSSION

Serbia comprises two main geochemical areas. One extends south of the river Danube and has large deposits of lead, zinc, antimony and copper. The other one extends north of the Danube (Vojvodina) and consists of quaternary sediments. For the genesis and development of agricultural soil in Vojvodina loess is the most widespread parental substrate. Loess provides the basis for the most important soil type in this region – chernozem. Loess in Vojvodina contains 20-30% of CaCO₃, which gives it an advantage over the loess in Eastern Europe (Živković, 1971). Such a favorable geological soil composition and pH is reflected in a more efficient selenium uptake by plants and a higher selenium concentration in plants (Fordyce *et al.*, 2000).

In the here described study the obtained results clearly indicate that the analyzed grain samples (n=68) collected on locations north of the Danube contained about twice as much selenium as the other geochemical areas. A similar observation was made by Klapac *et al.* (2004) regarding selenium content in vegetable samples collected in Croatia. The authors recorded a tendency of higher levels in samples from the river Sava basin, compared to the Drava basin, as the mean concentration of selenium was three times higher (114.2 ± 80.0 vs. 38.1 ± 1.0 µg/kg). Bratakos and Iannou (1989) tested 96 different locations in Greece and reported average selenium concentration in corn and wheat 120 ± 80 and 290 ± 190 µg/kg, respectively. Smrkolj *et al.* (2005) studied the selenium content in selected Slovenian foodstuffs, and reported an average selenium content of 11.9 µg/kg in wheat samples. Serdaru *et al.* (2003) examined the selenium status of 185 feeding stuff samples (hay, green plants and feedstuff concentrates) cultivated in south-west Romania. Only 6.5% of the samples contained an appropriate selenium content (150 – 300 µg/kg), while 93.5% of the samples were selenium deficient with a selenium content range of 1 – 150 µg/kg. Adams *et al.* (2002) conducted a survey on 452 grain samples of bread making wheat produced in the UK and reported a mean concentration of 32 µg/kg. Comparable selenium concentrations in grains were reported in other European countries (Tamás *et al.*, 2010). In general, European grains contain lower levels of selenium than North American samples. Very low selenium concentrations have been reported in Scandinavian countries with concentrations ranging 7 -18 µg/kg (Gissel – Nielsen, 1984).

Walnik *et al.* (1983) reported a range of 10-5300 µgSe/kg, with a mean value of 160 µg/kg for 290 wheat samples collected from major growing areas in USA. Comparable results were reported for Canada and Australia (Boila *et al.*, 1993; Lyons *et al.*, 2005).

All the above cited reports should be viewed under the light of nutritional requirements. The nutritional minimum level both for animals and humans is about 50 – 100 µgSe/kg in dry fodder/food, and intake below may cause selenium deficiency (Gissel – Nielsen, 1984). The National Research Council (NRC, 1985)

proposed the recommended minimal selenium concentrations in feedstuffs for sheep to be 100 µgSe/kg. The average selenium concentrations on each of the sampled locations south of the river Danube is below the minimum NRC recommended levels (Table 1). Tested grain samples grown in the Vojvodina region are marginally selenium deficient (mean concentrations ranging from 60.0 µg/kg in Melenci to 44.0 µg/kg in Vršac). The results obtained in our study are close to the results obtained by Šovljanski *et al.* (1991) who reported an average selenium concentration in wheat samples in Vojvodina in the range 50 - 60 µg/kg, and in Zaječar (36 µg/kg). The Sjenica plateau is dramatically selenium deficient with an average content of selenium in barley samples as low as 2 µg/kg (Mihailović *et al.*, 1996). These results being comparable to the values obtained in this study (4.3 µg/kg).

Based in extensive previous studies in China, Tan and Huang (1991) proposed the following ranges of grain selenium concentrations: below 25 µg/kg deficient, 25 – 40 µg/kg marginal, 40 -1000 µg/kg moderate to high, and above 1000 µg/kg excessive.

The absence of an existing pattern in selenium distribution in forage samples (Table 1) can be attributed to the heterogeneity of the botanical composition of the samples and differences in soil selenium uptake by plants. Pešut (1995) reported values for selenium concentration in alfalfa hay samples to be almost double compared to meadow hay sampled from the same region (70.79 ± 44.35 and 39.07 ± 18.20 µgSe/kg, respectively).

In Serbia in 2011 there was a total of 1.460.000 sheep, however the number is on the decrease as in 1991 a total of 2.127.000 sheep were recorded. The demand for meat is on the increase and production should be improved not only by increasing the number of animals, but by improving the zootechnical protocols, too. In order to determine the presence of nutritional selenium deficiency in sheep, and thus eventual losses due to unthriftiness and poor reproduction a survey on the selenium status of grazing sheep was carried out in our study.

The concentration of selenium in whole blood, plasma/serum and the liver of animals, as well as GPx activity is widely used by laboratories to predict the selenium status of animals (Gerloff, 1992). It is recognized that the information provided by whole blood selenium status represents an altogether different time frame in the nutritional history of the animal, as selenium is incorporated in the erythrocytes at the time of erythropoiesis (Nicholson *et al.*, 1991). Pavlata *et al.* (2012) assessed the correlation between the activity of GPx and the selenium concentrations in the whole blood of sheep, in order to calculate by linear regression analysis the activity of GPx corresponding to selenium concentrations between 70 and 100 µgSe/L whole blood. This interval is regarded as the reference value for sheep. The obtained results showed a high correlation between selenium status and GPx activity ($R^2=0.90$). The authors (Pavlata *et al.*, 2000; Scholz and Stober, 2002) previously described three stages of evaluation: adequate (higher than 100 µgSe/L whole blood) marginal, (70 – 100 µgSe/L) and deficient (less than 70

µgSe/L). The strongest correlation was found for selenium blood values below 70 µgSe/L, and the correlation between selenium concentration and GPx becomes less significant with higher selenium concentrations.

The results for GPx activity presented in Table 2 clearly show a pattern similar to the measured concentrations of selenium in grain samples. Regions with a higher selenium content in feedstuffs (north of the river Danube) reflected higher activities in whole blood GPx (Belo Blato 236.51 ± 85.72 µkat/L; Temerin 199.5 ± 83.24 µkat/L; Čoka 194 ± 101.07 µkat/L). However, these values are far lower than recommended by Pavlata *et al.* (2012) as the authors measured the whole blood GPx activity to be in the range 409 – 637 µkat/L for marginal selenium blood concentration (70 – 100 µgSe/L). Thus it can be concluded that the feedstuffs grown in the Vojvodina region cannot result in adequate blood selenium concentrations which would ensure an activity of GPx close to 600 µkat/L. Whole blood GPx activity measured on the other locations (Ub and Aleksinac) in Serbia are low (70.21 ± 20.15 µkat/L and 65.4 ± 11.9 µkat/L, respectively). Such values are in agreement with the report published by Hudman *et al.* (1988) that at GPx activities below 165 µkat/L deficiency signs, such as unthriftiness and nutritive muscular dystrophy (WMD) in lambs can be expected. The same group of authors described cases of WMD in cases where the GPx activity was 385 µkat/L, thus indicating to the need for additional prudence when estimating selenium deficiency only according to the presence of this disease.

ACKNOWLEDGEMENT

This work was supported by the Ministry of Science and Technology of Serbia, Grant No TR 31003 and Grant No TR 31050.

Address for correspondence:
Prof dr Olivera Valčić
Department of Physiology and Biochemistry
Faculty of Veterinary Medicine, University of Belgrade.
Bulevar Oslobođenja 18
11000 Belgrade, Serbia
Email: olja@vet.bg.ac.rs

REFERENCES

1. Adams ML, Lombi E, Zha FJ, McGrath SP, 2002, Evidence of low selenium concentrations in UK bread-making wheat brain, *J Sci Food Agric*, 82, 1160-5.
2. Boila RJ, Stothers SC, Campbell LD, 1993, The concentration of selenium in the grain from wheat, barley and oats grown at selected locations through Manitoba, *Can J Anim Sci*, 73, 217-21.
3. Bratakos MS, Ioannou PV, 1989, The regional distribution of selenium in Greek cereals, *Sci Total Environ*, 84, 237-47.
4. Brody T, 1994, Nutritional biochemistry, Academic Press, Inc: New York, NY.

5. Drake C, Grant AB, Hartley WH, 1960, Selenium in animal health, 2, The effect of selenium in unthrifty, weaned lambs, *N Z Vet J*, 8, 7-10.
6. Fordyce FM, Zhang GD, Green K, Liu XP, 2000, Soil, grain and water chemistry in relation to human selenium - responsive diseases in Enshi district, *Appl Geochem*, 15, 117-32.
7. Gerloff BJ, 1992, Effect of selenium supplementation on dairy cows, *J Anim Sci*, 70, 3934-40.
8. Gissel-Nielsen G, 1971, Influence of pH and texture of the soil on plant uptake of added selenium, *J Agr Food Chem*, 19, 1165.
9. Gissel-Nielsen G, 1974, Effect of fertilization on uptake of selenium by plants, In: Plant Analysis on Fertilizer Problems, Proceedings of the International Colloquium on Plant Analysis and Fertilizer Problems, Hanover, W Germany, Vol 1, 111.
10. Gissel-Nielsen G, Gupta UC, Lamand M, Westermarck T, 1984, Selenium in soils and plants and its importance in livestock and human nutrition, *Adv Agr*, 37, 397-460.
11. Günzler WA, Kremers H, Flohe L, 1974, An improved coupled test procedure for glutathione peroxidase (E.C. 1.11.1.9) in blood, *Z Klin Chem Klin Biochem*, 12, 444.
12. Hudman, JF, Costa ND, Robinson WF, 1988, An apparent phosphate selenium interaction in weaner sheep, *J Trace Elem Electrolytes Health Dis*, 2, 2, 105.
13. Klapac T, Mandić LM, Grgić J, Primorac Lj, Perl A, Krstanović V, 2004, Selenium in selected foods grown or purchased in eastern Croatia, *Food Chem*, 85, 445-52.
14. Kubota J, Allaway WH, Carter DL, Cary EE, Lazar VA, 1967, Selenium in crops in the United States in relation to selenium-responsive diseases of animals, *J Agr Food Chem*, 15, 3, 448-53.
15. Lindberg S, 1968, Selenium determination in plant and animal material, and water, A methodological study, *Acta Vet Scand Suppl*, 23, 1-41.
16. Liu CH, Lu ZH, Su Q, Duan YQ, 1986, Regional selenium deficiency of feeds in China, In: Proceedings of the Third International Symposium on Selenium in Biology and Medicine, Combs GFJ, Spallholz JE, Levander OA and Oldfield JE, Avi Publ Co, Westport, Conn.
17. Lyons GH, Genc Y, Stangoulis JCR, Palmer LT, Graham RD, 2005, Selenium distribution in weath grain, and the effect of postharvest processing on wheat selenium content, *Biol Trace Elem Res*, 103, 155-68.
18. Mihailović M, Kuzmanovski D, Črčev D, Jovanović I, Ledina O, Veličkovski S, 1993, Selenium status in cereals and selenium status of ewes and goats in Macedonia, 2 International summer conference for advancement of sheep and goat production, 100.
19. Mihailović M, Lindberg P, Jovanović IB, 1996, Selenium content in feedstuffs in Serbia, *Acta Vet* (Belgrade), 46, 5-6, 343-8.
20. Muth O, Oldfield J, Remmert L, Shubert J, 1958, Effects of selenium and vitamin E on white muscle disease, *Science*, 128, 1090.
21. National Research Council, 1985, Nutrient requirements of sheep, *National Academy Press*, 6th rev ed, Washington DC.
22. Nicholson JEG, Allen JG, Bush BS, 1991, Comparison of responses of whole blood and plasma selenium levels during selenium depletion and repletion of growing cattle, *Can J Anim Sci*, 71, 925 - 9.
23. Olson OE, Palmer IS, 1976, Selenoamino acids in tissues of rats administered inorganic selenium, *Metabolism*, 25, 299-306.
24. Osame S, Ohtani T, Itchio S, 1990, Studies on serum tocopherol and selenium levels and blood glutathione peroxidase activities in lambs with White Muscle Disease, *Jpn J Vet Sci*, 52, 705-10.
25. Pavlata L, Misurova L, Pechova A, Husakova T, Dvorak R, 2012, Direct and indirect assessment of selenium status in sheep – a comparison, *Vet Med*, 57, 5, 219-23.
26. Pavlata L, Pechova A, Illek J, 2000, Direct and indirect assessment of selenium status in cattle – a comparison, *Acta Vet Brno*, 69, 281 - 7.
27. Pešut O, 1995, Selenium content in feedstuffs and selenium status of sheep in the Bačka region, Master Thesis, University of Belgrade, Belgrade.

28. Rosenfeld I, Beath OA, 1964, Selenium: Geobotany, Biochemistry, Toxicity and Nutrition, Academic Press, New York.
29. Sankari S, Atroshi F, 1983, Effect of dietary selenium on erythrocyte glutathione peroxidase and blood selenium in two types of Finnisheep genetically selected for high and low glutathione peroxidase activity, *Zbl Vet Med A*, 30, 452.
30. Scholz RW, Hutchinso LJ, 1979, Distribution of glutathione peroxidase activity and selenium in the blood of dairy cows, *Am J Vet Rec*, 40, 2, 245.
31. Scholz H, Stober M, 2002, Enzootic myodystrophia in preruminant calves, In: Internal Medicine and Surgery in Cattle in German, Parey Buchverlag, Berlin, 1000-4.
32. Schwartz K, Folz CM, 1958, Factor 3 activity of selenium compounds, *J Biol Chem*, 233, 245.
33. Serdaru M, Vladescu L, Avram N, 2003, Monitoring of feed selenium in a southeast region of Romania, *J Agric Food Chem*, 51, 16, 4727-31.
34. Shepard, AD, Miller KR, 1981, Stability of GSH-Px in ovine blood samples under various storage conditions and response of this enzyme to different methods of selenium supplementation, *N Z Vet J*, 29, 77.
35. Smrkolj P, Pograjc L, Hlastan-Ribič C, Stibilj V, 2005, Selenium content in selected Slovenian feedstuffs and estimated daily intake of selenium, *Food Chem*, 90, 691-7.
36. Swaine J, 1955, The trace element content of soils, Commonwealth Bureau of Soil Sci Tech Comm, 48, 1-157.
37. Šovljanski R, Lazić S, Obradović S, Beker D, 1991, Sadržaj teških metala, selena i ostaci organohlornih insekticida u pšenici, *žito-hleb*, 18, 17.
38. Tamás M, Mándoki Zs, Csapó J, 2010, The role of selenium content of wheat in the human nutrition, A literature review, *Acta Univ Sapientiae, Alimentaria*, 3, 5-34.
39. Tan JN, Huang YJ, 1991, Selenium in geo-ecosystem and its relation to endemic diseases in China, *Water Air Soil Pollut*, 57-58, 59-68.
40. Terry N, Zajed AM, De Souza MP, Tarun AS, 2000, Selenium in higher plants, *Annu Rev Plant Physiol Mol Biol*, 51, 401 – 2.
41. Van Ryssen, Coertze RJ, Smith MF, 2013, Time-dependent effect of selenium supplementation on the relationship between selenium concentrations in whole blood and plasma of sheep, *Small Rum Res*, 112, 85-99.
42. Waldner C, Campbell J, Jim GK, Guichon PT, Booker C, 1998, Comparison of 3 methods of selenium assessment in cattle, *Can Vet J*, 39, 225-31.
43. Wilson PS, Judson GJ, 1976, Glutathione peroxidase activity in bovine and ovine erythrocytes in relation to blood selenium concentration, *Br Vet J*, 132, 428.
44. Wolnik KA, Fricke FL, Capar SG, Braude GL, Meyer MW, Satzger RD et al, 1983, Elements in major raw agricultural crops in the United States. 2. Other elements in lettuce, peanuts, potatoes, soybeans, sweet corn, and wheat, *J Agric Food Chem*, 31, 1224-49.
45. Živković B, Nejrgrbauer V, Tanasjević Đ, Milković N, Stojković L, Drezgic P, 1971, In: Zemljišta Vojvodine, Institut za poljoprivredna istraživanja, Novi Sad, 32-46.

STATUS SELENA U HRANIVIMA ZA OVCE NA ISPAŠI U SRBIJI

VALČIĆ OLIVERA, JOVANOVIĆ I, MILANOVIĆ SVETLANA I GVOZDIĆ D

SADRŽAJ

Ispitano je ukupno 221 uzoraka biljnih hraniva (187 žitarica i 34 uzoraka sena) sakupljenih sa različitih lokacija na području 15 opština Republike Srbije. Rezultati izraženi u $\mu\text{g}/\text{kg}$ pokazali su da je prosečan sadržaj selena 34.3 ± 17.1 $\mu\text{g}/\text{kg}$ u uzorcima žitarica i 53.8 ± 18.7 $\mu\text{g}/\text{kg}$ u uzorcima sena. Međutim, značajna razlika je uočena između uzoraka koji potiču sa područja severno (52.8 ± 20.0 i 73.4 ± 21.3 za uzorke žitarica i sena) u odnosu na uzorke sa južne strane toka Dunava (23.3 ± 15.4 i 41.7 ± 17.1 za uzorke žitarica i sena).

Aktivnost GPx je merena u ukupno 58 uzoraka krvne plazme ovaca koje su bile na ispaši na 5 različitih lokacija. Prosečna GPx aktivnost je iznosila 157.4 ± 61.9 $\mu\text{kat}/\text{L}$. Aktivnost GPx je pokazala distribuciju po lokalitetima slično kao i ispitivana biljna hraniva. Naime, aktivnost GPx je bila viša (212.8 ± 91.2 $\mu\text{kat}/\text{L}$) u uzorcima krvi ovaca na ispaši severno od Dunava u odnosu na ovce koje su bile na ispaši na južnim lokalitetima (66.9 ± 14.0 $\mu\text{kat}/\text{L}$).

U skladu sa postignutim rezultatima, kao i literaturnim podacima, Srbija može da se svrsta u seleno deficitarne regione, pri čemu lokaliteti južno od toka Dunava pokazuju više izražen deficit.